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# **Baird-Parker Agar Base**

### **DM095**

#### **Intended Use**

For the isolation and enumeration of coagulase positive staphylococci from food and other materials.

### **Contents**

See pack label.

### Formulation\*

Material:	Concentration in medium:		
Peptone mixture	12.0g/litre		
Yeast extract	3.0g/litre		
Sodium pyruvate	10.0g/litre		
Glycine	7.5g/litre		
Lithium chloride	5.0g/litre		
Agar	19.0g/litre		
Final pH: $6.8\pm0.2$			

### Storage and shelf life

All dehydrated culture media containers should be kept tightly closed and stored in a dry place at 10 to 25°C until the expiry date shown on the pack label.

# **Precautions**

For *in vitro* diagnostic use only. Observe approved hazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet (available on request or via MAST® website).

### Materials required but not provided

Standard microbiological supplies and equipment such as loops, MAST® selective supplements, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

# **Procedure**

- Refer to pack label for quantities and volumes required. Prepare MAST® Baird-Parker Agar Base (DM095) by suspending the powder in distilled or deionised water. For sachet packs, dissolve the entire contents of the sachet in the volume shown on the label.
- 2. Autoclave at 121°C (15 p.s.i.) for 15 minutes.
- Cool to 50°C and add 50ml of MAST® Baird-Parker Supplement, egg yolk tellurite emulsion (DM097S), to each litre of basal medium.
- 4. Mix thoroughly, pour culture plates (15 to 20ml per plate) and allow to set.
- 5. Prepared culture plates may be used immediately or stored in plastic bags at 2 to 8°C for up to one month before use.

- Macerate the food sample in 0.1% peptone water and make serial dilutions.
- 7. Spread 0.1 to 1.0ml volumes of the dilutions over the surface of a dried plate using a glass rod.
- 8. Incubate plates aerobically for 18 to 24 hours at 35 to 37°C, plates should be reincubated for a further 24 hours if no presumptive *Staphylococcus aureus* colonies appear.

### Interpretation of results

After incubation record growth of organisms. Typical characteristics to note include: colony size, colour and the presence or absence of a lecithinase halo (clear zone) around the colony.

# **Quality control**

Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate expected performance. Do not use the product if the result with the control organism is incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

Test Organisms	Growth	Colour	Lecithinase halos
Staphylococcus aureus ATCC® 25923	Good	Black	+
Staphylococcus epidermidis ATCC® 14990	Poor to good	Black	-
Escherichia coli ATCC® 25922	None	-	N/A
Bacillus subtilis ATCC® 6633	Poor to fair	Brown	-

### References

Bibliography available on request.